



A review of botanical, phytochemical, and pharmacological properties of *Alcea rosea* L.

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Abstract

Background and aims: This article was written to better introduce *Alcea rosea* as a valuable medicinal plant to the medical and pharmaceutical communities and the general public for industrial and traditional uses as a valuable medicinal plant.

Methods: The results were gathered from scientific papers and databases to find flavonoids, phenolic acids, amino acids, fatty acids, anthocyanins, polysaccharides, and pharmacological activities in *A. rosea*. Sixty-two articles were found, and 51 were used for our final results.

Results: Studies have shown that flowers, roots, seeds, and other parts of the plant exert antioxidant, antimicrobial, anti-inflammatory, cytotoxic, sedative, analgesic and anti-cough effects due to the presence of valuable phytochemical compounds such as anthocyanins, pectin, starch, monosaccharide, disaccharide, mucilage, flavonoids (hypolaetin-8-glucoside, isoquercitrin, kaempferol, caffeic acid, p-coumaric acid), coumarins, scopolamine, phytosterol, tannins, asparagine, and certain amino acids.

Conclusion: *Alcea rosea*, as a rich source of secondary metabolites, can have diverse applications in medical and food research.

Keywords: *Alcea rosea*, Botany, Morphology, Pharmacological properties, Phytochemistry

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Introduction

Most of the information available on medicinal plants has been practically obtained from experiences with traditional uses (1). Medicinal plants contain substances with medicinal properties that may be used directly or as raw materials for the synthesis of commercially valuable drugs (2). Researchers have recently sought to replace chemical drugs with nature-based medications to prevent many drug-related problems. However, medicinal plants also have chemicals that, as with synthetic drugs, have the potential to cause relatively severe adverse side effects (2), so identifying the bioactive ingredients of medicinal plants and investigating their effects and the unpredictable effects of these compounds can help determine the use of these plants (1,3). Humans have used *A. rosea* for pharmaceutical purposes since 6000 years ago (4). In recent years, studies have shown that flowers, roots, seeds, and other parts of the plant exert antioxidant, antimicrobial, anti-inflammatory, cytotoxic, sedative, analgesic, anti-cough, immunomodulatory, and antibiotic effects (5) due to the presence of valuable phytochemical compounds such as anthocyanins, pectin, starch, monosaccharide, disaccharide, mucilage, flavonoids (hypolaetin-8-glucoside, isoquercitrin, kaempferol, caffeic acid, p-coumaric acid), coumarins,

scopolamine, phytosterol, tannins, asparagine, and certain amino acids (6-8). Therefore, *Alcea rosea*, as a rich source of secondary metabolites, can have diverse applications in medical and food research. This study aimed to investigate morphological characteristics, physicochemical properties, phytochemical compounds (phenol, flavonoids, fatty acids, proteins, and polysaccharides), and therapeutic properties of *A. rosea* to identify its properties and help conduct subsequent studies.

Materials and Methods

Information from herbal books, authoritative journals, and articles retrieved from databases such as Google Scholar, PubMed, Science Direct, and Scopus search engine using keywords, medicinal activities, physicochemical properties, traditional medicine, modern medicine, phytochemical compounds, and commercial products were collected in *Alcea rosea*. And then, after reviewing the primary articles, more detailed searches were done again, for example, in phytochemical compounds, the words flavonoids, phenolic acid, amino acid, fatty acid, anthocyanins, polysaccharides, and in modern medicine, the words Anticancer Activity, Antidiabetic Activity, Kidney Stone, Antioxidant Activity, Antimicrobial Effect, Immune System, Antiestrogenic Effects were searched

to get complete information. The text of the abstracts of the articles and the titles of the chapters of the books were checked in terms of thematic connection with the items mentioned above, and then the unrelated items were removed from among them. In the initial search, 62 cases were found, and after reviewing the findings in the final analysis, 51 of them were used. Then the report studied and used the details of articles, chapters, and different parts of flora and books.

Results

Classification, geographical distribution, botanical characteristics, and reproduction methods

Classification and local names of *A. rosea* in some

countries are presented in Table 1. This plant spreads worldwide, including the western Mediterranean region, to the center of Asia. This species is cultivated as an ornamental and medicinal plant or grows wild as a weed on the city's outskirts and the side of roads (9,10). Sunny regions are suitable for plant growth (11). The form of the plant is erect with few branches (12,13), it is 0.5 to 2 meters high, and its stem diameter is 2.4 to 3 cm. The leaves are large (7.5 to 12.5 cm in diameter) with 5-7 deep lobes. The flowers (Figure 1) are more significant than 10 cm in diameter, have different colors, and are in the Racom's inflorescence (12). The seed is kidney-shaped and disc-like, its edges are trichomes, and its size is 6 mm. The color of the seeds is brown to black and white, which

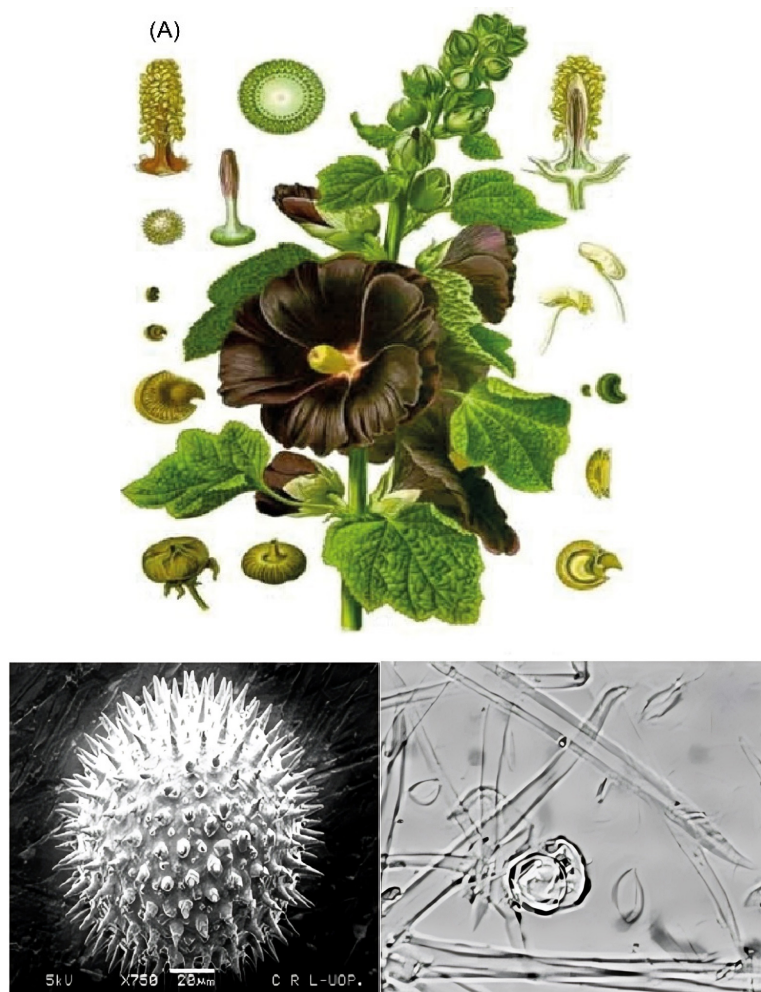


Figure 1. A: *A. rosea*. (16), B: Pollen grain of *A. rosea* (15), C: Peltate glands (type-II) on the abaxial surface of *A. rosea*

Table 1. Classification and local names of *A. rosea* in some countries

Vernacular names (6)		Plant taxonomy (14)	
Malta: Hollyhock, Malvarose	Arabic: Khatma wardi	Kingdom:	Plantae plants
Roumanian: Nalba de gardina	Chinese: Shu k'ui	Division:	Magnoliopsida
Russian: Chernaya roja, Shtok rosa	English: Hock Herb, Hollyhock, Round Dock	Class:	Magnoliopsida
Spanish: Malva arborea, Malva loca	French: Alcee, Alcee rose, Althee rose, Mauve rose, Pass rose	Ordea:	Malvaceae
Kannad: Doddabindigaegidda	German: Augenpappel, Baummalve, Baumrose, Gartenmalve, Rosenpappel	Genus:	<i>Alcea rosea</i> Linn
Italian: Malvarose, Malvoni	Greek: Altaia	Synonyms:	<i>Althaea rosea</i> Linn*

**Althaea rosea* (L.) Cav. is a synonym of *Alcea rosea* L. The record derives from WCSP (in review) (data supplied on 2012-03-23), which reports it as a synonym with original publication details: Diss. 2: 91 1786 (IPNI: <http://ipni.org/urn:lsid:ipni.org:names:558713-1>).

becomes distinct when the seed ripens. The pollen grains are spherical with a diameter of about 115 (132) 142 μm . At the pollen grains' surface, some stomata are at a pore size of 2 (4) 5 μm (14). In their study in 2010, Shaheen et al. observed that the maximum size of *A. rosea* pollen grain is 125 (143.2 \pm 4.2) 162.5 μm , and most stomata are diacytic, stellate trichomes and monomorphic (15).

Reproduction of this plant is often done through seed, but the division of the root is also used ("The Wealth of India," 1985); another way of reproducing this plant is tissue culture. Among the explants, cotyledons are more appropriate for callosing and organogenesis of *A. rosea* (17-19).

Physicochemical characteristics

The results of a study on *A. rosea* seeds showed that

moisture content was (8.2 \pm 0.38%) and inflation index (5.3 \pm 0.16) ml. Total ash, acid-insoluble, and water-soluble ash were 7.3 \pm 0.32, 1.48 \pm 0.16, and 3.33 \pm 0.24, respectively (20).

Phytochemical evaluation

Phytochemical compounds are secondary plant metabolites that may not be necessary for the plant but have essential properties, such as antioxidant (21), antimicrobial, antifungal (22,23), anti-inflammatory, and antidiabetic properties that are also used to prevent and treat diseases (9,24). These compounds of *A. rosea* are rich in alkaloids, carbohydrates, fatty acids, phenolic compounds, glycosides, flavonoids (25), metals, amino acids, and monosaccharides (26). The secondary metabolites reported to exist in *A. rosea* are shown in Table

Table 2. Secondary metabolites in *A. rosea*

Group	Compound	Reference
Amino acid	Valine, Threonine, Methionine, Isoleucine, Leucine, Lysine, Phenylalanine, Histidine, Arginine	(27)
	Linoleic acid	(28)
Fatty acid	Palmitic acid, Stearic acid, Oleic acid, Linolenic acid, Linoleic acid, Mirstin	(6,29-32)
	α -Terpenyl-OAc, Stigmasterol, <i>p</i> -Tolualdehyde, β -Sitosterol	(6,29,30)
Phenolic acid	Seringic, <i>p</i> -Hydroxybenzoic, <i>p</i> -Comaric, Ferrolic acid, Caffeic, Vanillic, <i>p</i> -Hydroxy-phenylacetic acid, <i>m</i> -Hydroxybenzoic acids	(33,34)
	Chlorogenic	(33)
	Salicylic acid	(34)
Flavonoids	Kaempferol-3- <i>O</i> - β -D-glucopyranoside	(35)
	Kaempferol	(6,12,35-38)
	Quercetin	(6,12,28,29,33,37,38)
	Luteolin	(33)
	Quercetin-3- <i>O</i> - β -D-glucuronopyranoside-8-C- β -D- glucopyranoside, 3- <i>O</i> - β -D-rutinoside, Kaempferol -4'- <i>O</i> - β -D-glucoside, Kaempferol -3 - <i>O</i> - β -D-glucoside.	(35)
	Dihydrokaempferol-4- <i>O</i> -glucopyranoside, Dihydrokaempferol-4'- <i>O</i> - β -D-glucopyranoside, dihydrokaempferol, Kaempferol-3- <i>O</i> -[6''-(<i>E</i> -coumaroyl)]- β -D-glucopyranoside, Kaempferol-3- <i>O</i> - β -D-glucopyranoside, Apigenin, Kaempferol-3- <i>O</i> - α -L-rhamnopyranosyl-(1'' \rightarrow 6'')- β -dglucopyranoside	(39)
	Isoquercetin	(12,28,29)
	Kaempferol-3- <i>O</i> -glucoside	
	Trigonelline	(40)
	Glycine betaine	
Coumarin	(6,12,37,38)	
Anthocyanins	Cyanidine	(12,28,29)
	Delphinidin-3- <i>O</i> -glucoside, Delphinidin-3- <i>O</i> -rutinoside, Cyanidin-3- <i>O</i> -glucoside, Cyanidin-3- <i>O</i> -rutinoside, Petunidin 3- <i>O</i> -glucoside, Petunidin-3- <i>O</i> -rhamnopyranosylglucoside, Malvidin- 3- <i>O</i> -glucoside, Malvidin-3- <i>O</i> -malonylglucoside, Malvidin-3- <i>O</i> -rhamnopyranosylglucoside, Myricetin-3- <i>O</i> -glucoside, Kaempferol 3- <i>O</i> -rutinoside, Kaempferol-3- <i>O</i> -glucoside, Luteolin-4'- <i>O</i> -glucoside	(41)
	Dolphinidine, Petotnidine, Malvidine, Dolphinidine, Phetodine and Malvydine from (3- <i>O</i> -glucoside), Dolphinidine, Penthidine and Malvydine from (3,5-di- <i>O</i> -glycosides)	(42)
	Luteolin 8-C-glucoside, 7-methyl ethers	(43)
Polysaccharides	Galactronic Acid, Glucuronic Acid, Ramenose, Arabinose, Galactose	(26,39,44-46)
	Glucose	(26,44,47)
	Mannose, xylitol	(44,47)
	Cyclohexanol, Limonene, Phellandrene, β -Sitosterol, besides sucrose, <i>p</i> -Tolualdehyde, α -terpenyl acetate	(45)
	Starch, Lignin	(28, 48)
	Galacturonorhamnans, Arabinans, Glucans, Arabinogalactans, Glucans	(6, 12, 37, 38)
Hemicellulose, Uranic acid	(26)	

2, and the schematic structure of some of its compounds is in Table 3.

Pharmacological activities

Therapeutic uses in traditional medicine

In different countries of the world (Ecuador, Saudi Arabia, India, Italy, etc), decoction, infusion, crude, ointment of the flower, root, stem, and seeds of *A. rosea* are traditionally used in the treatment of cough, asthma, throat infection, dysuria, rash, kidney stones and pain,

Table 3. The schematic structure of some of the compounds that have been reported to exist in *A. rosea*

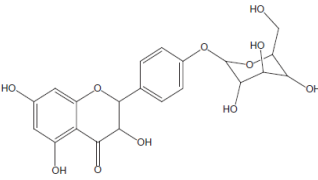
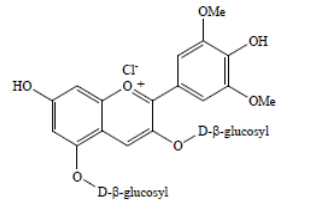
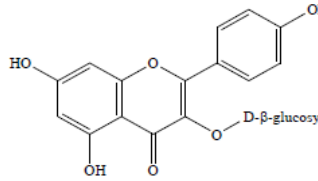
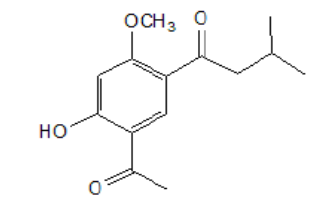
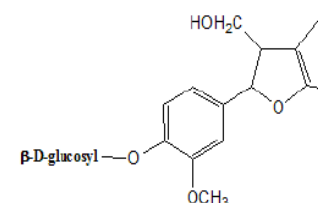
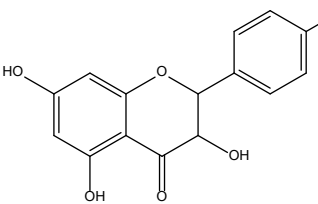
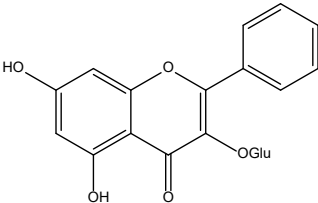
Molecular Structure	Isolated Compound	Reference
	Dihydrokaempferol-4'-O glucopyranoside	(39)
	Malvidin-3,5-diglucoside	(49)
	Astragalin	(49)
	Rosea B	(49)
	Rosea A	(49)
	Dihydrokaempferol-4'-O-beta-dglucopyranoside	(39)
	Dihydrokaempferol	(39)

Table 3. Continued.

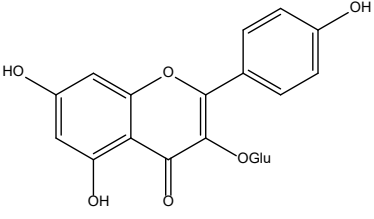
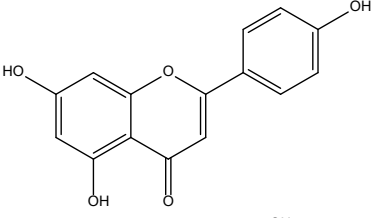
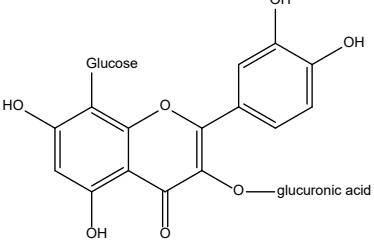
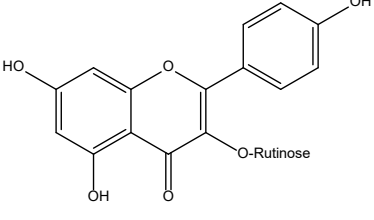
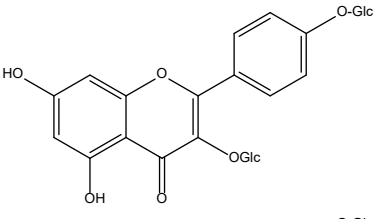
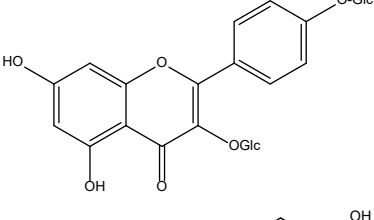
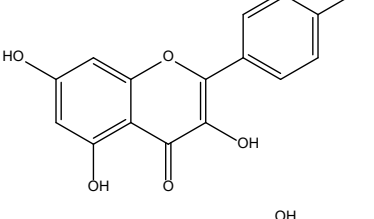
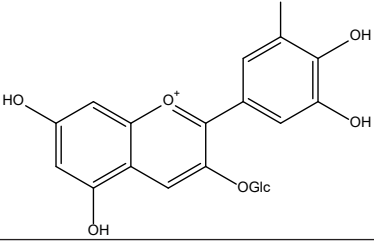
	Kaempferol-3- <i>O</i> - β -D-glucopyranoside	(39)
	Apigenin	(39)
	Quercetin-3- <i>O</i> - β -D-glucuronopyranoside-8- <i>C</i> - β -D-glucopyranoside	(35)
	Kaempferol-3- <i>O</i> - β -D-rutinoside	(35)
	Kaempferol-4'- <i>O</i> - β -D-glucoside	(35)
	Kaempferol-3- <i>O</i> - β -D-glucoside	(35)
	Kaempferol	(35)
	Delphinidin-3- <i>O</i> -glucoside	(41)

Table 3. Continued.

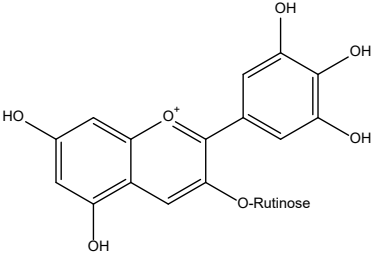
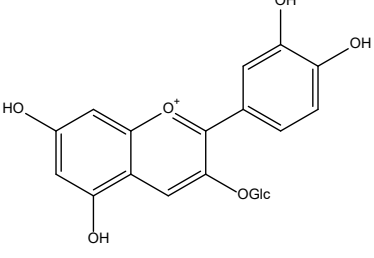
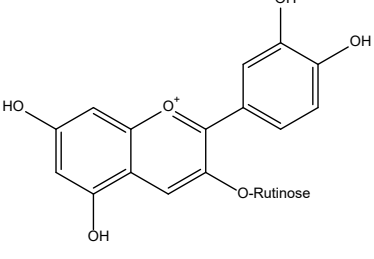
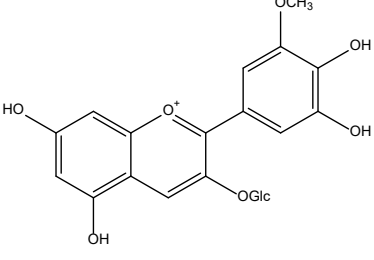
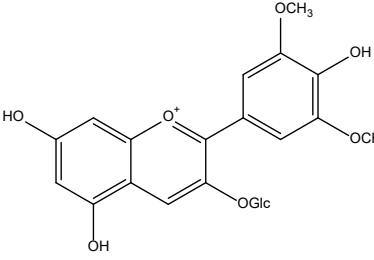
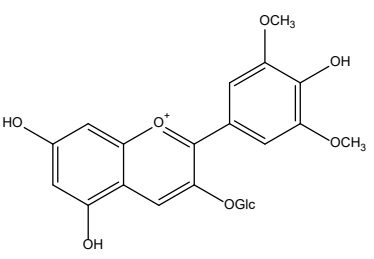
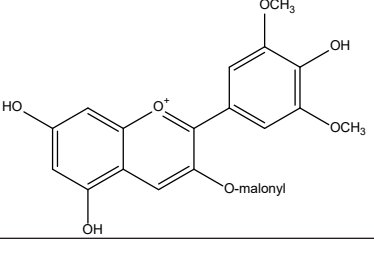
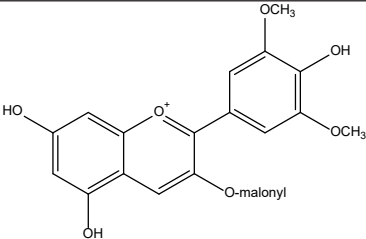
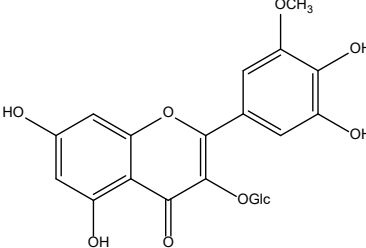
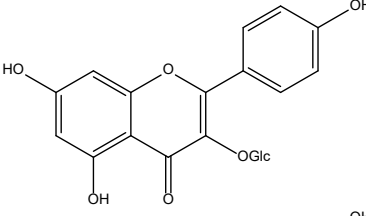
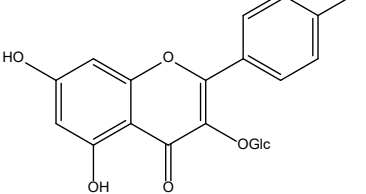
	Delphinidin 3-O-rutinoside	(41)
	Cyanidin 3-O-glucoside	(41)
	Cyanidin 3-O-rutinoside	(41)
	Petunidin 3-O-glucoside	(41)
	Petunidin 3-O-rhamnosylglucoside	(41)
	Malvidin 3-O-glucoside	(41)
	Malvidin 3-O-rhamnosylglucoside	(41)

Table 3. Continued.

	Malvidin 3-O-malonylglucoside.	(41)
	Myricetin 3-O-glucoside	(41)
	Kaempferol 3-O-rutinoside	(41)
	Kaempferol 3-O-glucoside	(41)

and jaundice (50,51). This plant is a diuretic, refrigerant, sedative, softener, antipyretic, and astringent and is used for wound healing, bronchitis, reducing appetite, and angina, analgesic, arthritis, dysuria (6,12,52-56). It is also used to improve blood circulation, constipation, reduce menstrual pain and bleeding (57), goiter, dermatitis, and liver diseases, mastitis, insect bites (50,51,57-59). In Iran, decoction, infusion, crude, ointment, and parts of *A. rosea* are traditionally used to treat digestive, respiratory, skin, cold, and migraine problems (60,61).

Therapeutic uses in modern medicine

Anticancer activity

The methanolic extract of the seed, root, and whole plant, in vivo conditions, has been reported to exert significant anticancer properties that control neoplastic cell changes in some of the cancer cell lines, such as JB6 + of the epidermis, in condition vivo (5,55,62,63). The ethyl acetate extract of *A. rosea* seed has been reported to inhibit proliferation and colony formation dose-time-dependently and to promote apoptosis, as confirmed by PARP cleavage and an increase in Bax expression, along with a reduction in BCL-xL protein levels in HCT116 and SW480 cells, respectively. In addition, *A. rosea* seed extract causes cell cycle arrest at the G0/G1 phase and

decreases cyclin D1. The ethyl acetate extract of *A. rosea* seed treatment also reduced the number and size of colon spheres in the cells dose-dependently and simultaneously decreased the ALDH1A1 and Dclk1 cancer stem cell markers. After treatment with *A. rosea* extract, relative levels of -catenin, Notch-ICD, Hes1, and EZH2 decreased. TOP-flash reporter activity used to measure Wnt signaling declined significantly after treatment, while overexpression of wild-type but not mutant EZH2 neutralized the inhibitory effects. In addition, WIF1 (a Wnt antagonist) promoter activity increased notably after *A. rosea* extract treatment, phenocopying an increase in WIF1 reporter activity after EZH2 knockdown. *A. rosea* extract inhibited tumor growth in vivo, most likely by lowering levels of EZH2, -catenin, cyclin D1, and Ki-67, as well as cancer stem cell markers (62). Methanolic extract of *A. rosea* root in vivo has been observed to significantly suppress neoplastic cell transformation by inhibiting EGF receptor (EGFR) kinase activity. After treatment with methanolic *A. rosea* extract root, EGFR activation by EGF was inhibited in EGFR +/+ cells but not in EGFR -/- cells. In addition, MARC inhibited EGF-induced cell proliferation in EGFR-expressing murine embryonic fibroblasts (EGFR +/+) (5).

Antidiabetic activity

The *A. rosea* seed has shown antihyperglycaemic and antioxidant activities in rats with alloxan-induced diabetes. Methanolic and aqueous extracts at 300 µg/kg reduce blood glucose levels and increase antioxidant levels in the liver tissue (59). Some other researchers have also reported the plant as an antidiabetic agent (64, 65).

Kidney stone treatment

The hydroalcoholic extract of this plant significantly reduces the calcium oxalate deposits of the kidney and also decreases the urinary oxalate levels in the rats induced by ethylene glycol; in other words, it can be widely used to treat kidney stones due to its anti-urolithiasis and diuretic properties (55).

Cardiovascular effects

Alcoholic extract of *A. rosea* L. flower increased the outflow of the coronary artery of the heart isolated from guinea pigs. It pronouncedly dilated the blood vessels in the rat's hind limbs. The extract produced a temporary hypotensive effect on anesthetic cats. It inhibited ADP-induced platelet aggregation and exerted an inhibitory effect on experimental thrombosis formation (66).

Hepatoprotective effects against acetaminophen-induced hepatotoxicity

A histopathologic study of liver tissues showed that *A. rosea* extracts produced protective effects against acetaminophen-induced hepatic toxicity (67).

Anti-estrogenic effects

Berger et al (68) and Dudek et al (33) reported that cinnamic acid derivatives, para-coumaric acid, ferulic acid, and p-hydroxybenzoic acid were found throughout *A. rosea* and had estrogenic activity. Researchers have also reported weak estrogenic activity in rat testis for the aqueous *A. rosea* extract (69). The latter study studied a methanolic extract containing more flavonoids than the aqueous one. The morphological and histo-enzymatic changes in the Leydig cells indicated that the methanolic *A. rosea* extract produced a direct but partial effect on rat testis. The insignificant changes in estradiol content and testicular testosterone suggest that the extract does not impair steroidogenesis (70).

Effect on the immune system

One study has confirmed the immunogenetic activity (64). Another study showed that aqueous extract of *A. rosea* produces the following effects on the immune system: Inducing a transient non-specific polyclonal response evidenced by the production of IL-4 in non-immunized, treated mice; initially boosting the output of anti-EA antibodies and IL-4, a T-helper two cytokines; and suppressing the production of gamma-interferon, a T-helper 1 cytokine (71).

Antioxidant activity

Free radicals cause inflammation, cancer, cardiovascular disease, and osteoporosis; therefore, antioxidant compounds benefit human health and reduce oxidative stress. The antioxidant activity of *A. rosea* has already been reported (5, 35, 62, 72, 73). The antioxidant activity of methanolic extracts of aerial parts and flowers of *A. rosea* has been measured more frequently than that of the vegetative part (herbage). In the investigations of Liu et al (73), the antioxidant activity of aqueous, ethanol, butanol, and chloroform extracts *A. rosea* seed was measured using four superoxide anion scavenging, hydroxyl radical scavenging assay, superoxide dismutase activity assay, and DPPH assay. The aqueous extract had the most significant antioxidant activity in the DPPH method, superoxide dismutase activity assay, and superoxide anion scavenging assay. Butanol extract had comparatively more significant antioxidant activity in the hydroxyl radical scavenging assay. Antioxidant activity was directly related to the total phenolic content, so the h distilled water extracts found the highest total phenolic content. The chloroform extract exhibited the weakest antioxidant activity (73). Tyrosinase activity was reported to be inhibited by hydroalcoholic *A. rosea* extract (74). Differences in antioxidant activity are due to differences in the amounts of secondary metabolites, such as flavonoids, or the presence of other secondary antioxidant compounds that directly or indirectly produce this activity (75).

Antimicrobial Effect

Several studies have confirmed the antimicrobial effects (Tables 4 and 5) of this plant's different extracts (64, 76).

Other properties

The ethyl acetate extract showed cytotoxic activity against Brine shrimp (LC₅₀ < 1000) (77), but this extract was even more active than umbelliferon (78) and colchicine (79). Choi et al (5) studied whether methanol extracts of *A. rosea* were cytotoxic in the mouse epidermal JB6 P + cells. Their results showed that methanol *A. rosea* extract at different doses did not affect the viability of JB6 P + cells (5). In addition, the compound kaempferol-3-O-[6''-(E-coumaroyl)]-β-d-glucopyranoside extracted from *A. rosea* has a potent cytotoxic activity on HepG-2 cell line with an IC₅₀ of 3.822 µg/mL (6.434 µM) when compared to reference standard 5-Fluorouracil (IC₅₀ = 0.9 µg/mL, 6.919 µM) (63). Methanol and aqueous extracts of *A. rosea* seed showed nontoxic effects on the behavioral parameters in normal healthy rats. A survival rate of 100% was observed in animals treated with 50–300 mg/kg body weight (BW) of *A. rosea* extracts, while in the animals treated with the extracts at 350–500 mg/kg BW, the survival rate was 50–90% (59). The dry extract of this plant stem and root has been found to contain monosaccharides, oligosaccharides, mucus, microelements, and proteins. Therefore, the dry extract of this plant has therapeutic effects on asthma, bronchitis, respiratory tract infection, and inflammatory

disease (26).

Commercial products

Alcea rosea makes paper bags (80) and edible colors (71). So far, no pharmaceutical and cosmetic products have been reported to be produced from the *Alcea* genus.

Discussion and Future Perspectives

Results have shown that *A. rosea* is one of the most valuable medicinal plants with critical medicinal compounds such as mucilage and high phenolic compounds. This plant grows wild in Iran in abundance, which the general public traditionally collects to treat diseases. In addition, this plant is cultivated commercially and has been used to make many medicines. Hence, the economic significance of the medicinal *A. rosea* is undeniable. Due to the potential, and diverse applications of active ingredients of medicinal *A. rosea*, especially its medicinal compounds, their use can reduce the use of synthetic drugs that threaten the immunity of consumers. Hence, the importance of using the active ingredients of this plant multiplies the value of research related to this issue and prompts further

research on the other valuable and unknown properties of this plant to be used as a medicine for treating human diseases. The key that can be used in this research is the anti-inflammatory, analgesic, antifungal, and therapeutic effects for digestive, skin, respiratory, and other medicinal diseases of *A. rosea*, which is due to the presence of valuable medicinal compounds such as mucilage, which has been less investigated. It is also recommended to use this plant to develop cosmetic-medicinal products due to its potential.

Conclusion

This research data will help you get to know this plant, its medicinal properties, and other uses in the medical, food, agricultural, etc. industries. Our research showed that *A. rosea* is one of the most important and widely used medicinal plants because it is used in the design of green spaces and gardens due to its beautiful appearance and also because of its valuable compounds; the traditional and modern uses of this plant and its therapeutic effects have been reported in various diseases. The information

Table 4. Antibacterial effects of different concentrations of *Alcea rosea* ethanol extract (80)

Bacterial specie	Different concentrations of <i>A. rosea</i> ethanol extract							
	Flower				Leaf			
	0.05	0.10	0.20	0.40	0.05	0.10	0.20	0.40
Gram-positive bacteria								
<i>Bacillus anthracis</i>	7	10	11	15	8	9	9	14
<i>Bacillus cereus</i>	R	R	R	9	R	R	8	10
<i>Staphylococcus aureus</i>	10	13	17	18	8	8	10	13
<i>Staphylococcus epidermidis</i>	9	10	15	19	R	9	10	13
<i>Listeria monocytogenes</i>	R	R	R	7	R	R	7	10
<i>Streptococcus pyogenes</i>	8	9	10	13	8	8	9	11
Gram-negative bacteria								
<i>Escherichia coli</i>	R	R	R	R	R	R	R	R
<i>Salmonella Typhi</i>	R	R	7	8	R	R	R	8
<i>Klebsiella pneumoniae</i>	7	7	9	14	7	8	10	12
<i>Pseudomonas aeruginosa</i>	R	8	9	13	R	R	R	R

R: Resistant, *(6 mm) diameter disc.

Table 5. Antimicrobial activity of *Alcea rosea* extracts (81)

Microorganisms	Inhibition zone (mm)*							
	A	B	C	D	E	F	G	H
<i>Escherichia coli</i> ATCC 29998	9	9	9	9	9	15	-	-
<i>Escherichia coli</i> ATCC 25922	-	-	-	-	-	14	-	-
<i>Escherichia coli</i> ATCC11230	10	9	10	9	9	18	-	-
<i>Staphylococcus aureus</i> ATCC 6538P	8	10	9	7	9	12	-	-
<i>Staphylococcus aureus</i> ATCC 29213	-	9	9	9	9	13	-	-
<i>Staphylococcus epidermidis</i> ATCC 1222	8	10	10	9	8	12	-	-
<i>Salmonella Typhimurium</i> CCM 5445	8	8	8	8	7	14	-	-
<i>Enterobacter cloacae</i> ATCC 13047	-	-	-	-	-	13	-	-
<i>Enterococcus faecalis</i> ATCC 29212	-	-	-	-	-	11	-	-
<i>Pseudomonas aeruginosa</i> ATCC 27853	9	9	9	8	8	22	-	-
<i>Candida albicans</i> ATCC 10239	-	-	-	-	-	-	18	-

A: Ethanol extract; B: n-Hexane extract; C: Ethyl acetate extract; D: Methanol extract; E: Water extract; F: Ceftazidime; G: Nystatin; H: Control (DMSO).

*Includes the diameter of the disc (6 mm).

collected in this article can be a basis for further studies and help make herbal and natural medicines. In addition to knowing the importance of this medicinal plant, the necessity of cultivation and production of this medicinal plant and its secondary metabolites multiplies, which should be considered in the future.

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Competing Interests

Although one of the authors in this article is the journal's Editor-in-Chief, the whole process of reviewing and publishing this article is like other articles in the journal, and there is no difference in its review from other articles.

Ethical Approval

Not applicable.

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References

- Taylor JL, Rabe T, McGaw LJ, Jäger AK, van Staden J. Towards the scientific validation of traditional medicinal plants. *Plant Growth Regul.* 2001;34(1):23-37. doi: [10.1023/A:1013310809275](https://doi.org/10.1023/A:1013310809275).
- Jamshidi-Kia F, Lorigooini Z, Amini-Khoei H. Medicinal plants: past history and future perspective. *J Herbmed Pharmacol.* 2018;7(1):1-7. doi: [10.15171/jhp.2018.01](https://doi.org/10.15171/jhp.2018.01).
- Banaee M, Sureada A, Mirvaghefi AR, Rafei GR. Effects of long-term silymarin oral supplementation on the blood biochemical profile of rainbow trout (*Oncorhynchus mykiss*). *Fish Physiol Biochem.* 2011;37(4):885-96. doi: [10.1007/s10695-011-9486-z](https://doi.org/10.1007/s10695-011-9486-z).
- Cowan MM. Plant products as antimicrobial agents. *Clin Microbiol Rev.* 1999;12(4):564-82. doi: [10.1128/cmr.12.4.564](https://doi.org/10.1128/cmr.12.4.564).
- Choi ES, Cho SD, Shin JA, Kwon KH, Cho NP, Shim JH. *Althaea rosea* Cavanil and *Plantago major* L. suppress neoplastic cell transformation through the inhibition of epidermal growth factor receptor kinase. *Mol Med Rep.* 2012;6(4):843-7. doi: [10.3892/mmr.2012.977](https://doi.org/10.3892/mmr.2012.977).
- Chopra RN. *Chopra's Indigenous Drugs of India*. Calcutta: UN Dhur & Sons Pvt Ltd; 1958.
- Chopra RN, Chopra IC, Handa KL, Kapoor LD. *Indigenous Drugs of India*. 2nd ed. Calcutta: UN Dhur & Sons Pvt Ltd; 1958.
- Pullaiah T. *Encyclopaedia of World Medicinal Plants*. Vol 2. New Delhi: Regency Publications; 2006. p. 2442.
- Singh MP, Panda H. *Medicinal Herbs with Their Formulations*. New Delhi: Daya Publishing House; 2005. p. 73.
- Pakravan M. A new species and a new combination in Iranian *Alcea* (Malvaceae). *Ann Bot Fenn.* 2008;45(2):133-6. doi: [10.5735/085.045.0207](https://doi.org/10.5735/085.045.0207).
- Al-Snafi AE. The pharmaceutical importance of *Althaea officinalis* and *Althaea rosea*: a review. *Int J Pharmtech Res.* 2013;5(3):1387-5.
- Kirtikar KR, Basu BD. *Indian Medicinal Plants*. 2nd ed. Vol 1. Dehra Dun: International Book Distributors; 1987.
- Naqshi AR, Dar GH, Javeid GN, Kachroo P. Malvaceae of Jammu and Kashmir State, India. *Ann Mo Bot Gard.* 1988;75(4):1499-524. doi: [10.2307/2399298](https://doi.org/10.2307/2399298).
- Fahamiya N, Shiffa M, Aslam M. A comprehensive review on *Althaea rosea* Linn. *Indo Am J Pharm Res.* 2016;6(11):6888-94.
- Shaheen NI, Khan MA, Yasmin G, Hayat MQ, Munsif SH, Ahmad KH. Foliar epidermal anatomy and pollen morphology of the genera *Alcea* and *Althaea* (Malvaceae) from Pakistan. *Int J Agric Biol.* 2010;12(3):329-34.
- Hollyhock, *Alcea rosea* (*Althaea rosea*). Chromolithograph after a botanical illustration by Walther Muller from Hermann Adolph Koehler's *Medicinal Plants*, edited by Gustav Pabst, Koehler, Germany, 1887.
- Sakhanokho HF, Zipf A, Rajasekaran K, Saha S, Sharma GC. Induction of highly embryogenic calli and plant regeneration in upland (*Gossypium hirsutum* L.) and Pima (*Gossypium barbadense* L.) cottons. *Crop Sci.* 2001;41(4):1235-40. doi: [10.2135/cropsci2001.4141235x](https://doi.org/10.2135/cropsci2001.4141235x).
- Triplett BA, Moss SC, Bland JM, Dowd MK. Induction of hairy root cultures from *Gossypium hirsutum* and *Gossypium barbadense* to produce gossypol and related compounds. *In Vitro Cell Dev Biol Plant.* 2008;44(6):508-17. doi: [10.1007/s11627-008-9141-2](https://doi.org/10.1007/s11627-008-9141-2).
- Sylvère Sié R, Charles G, Sakhanokho HF, Toueix Y, Dje Y, Sangaré A, Branchard M. Protocols for callus and somatic embryo initiation for *Hibiscus sabdariffa* L. (Malvaceae): influence of explant type, sugar, and plant growth regulators. *Aust J Crop Sci.* 2010;4(2):98-106.
- Fahamiya N, Aslam M, Siddiqui A, Shiffa M, Husain A, Ahmad S, et al. Pharmacognostical, physicochemical and phytochemical investigation of *Althaea rosea* Linn. *Int J Pharm Res Dev.* 2011;4(3):129-40.
- Wong S, Lim Y, Chan E. Antioxidant properties of *Hibiscus*: species variation, altitudinal change, coastal influence and floral colour change. *J Trop For Sci.* 2009;21(4):307-15.
- Khan M, Wassilew SW, Schmutterer H, Ascher KR. *Natural Pesticides from the Neem Tree and Other Tropical Plants*. Eschborn, Germany: GTZ; 1987. p. 645-50.
- Nair R, Kalariya T, Chanda S. Antibacterial activity of some selected Indian medicinal flora. *Turk J Biol.* 2005;29(1):41-7.
- Nazarian-Samani Z, Sewell RD, Lorigooini Z, Rafieian-Kopaei M. Medicinal plants with multiple effects on diabetes mellitus and its complications: a systematic review. *Curr Diab Rep.* 2018;18(10):72. doi: [10.1007/s11892-018-1042-0](https://doi.org/10.1007/s11892-018-1042-0).
- Sadighara P, Gharibi S, Moghadam Jafari A, Jahed Khaniki G, Salari S. The antioxidant and flavonoids contents of *Althaea officinalis* L. flowers based on their color. *Avicenna J Phytomed.* 2012;2(3):113-7.
- Azizov UM, Mirakilova DB, Umarova NT, Salikhov SA, Rakhimov DA, Mezhlumyan LG. Chemical composition of dry extracts from *Alcea rosea*. *Chem Nat Compd.* 2007;43(5):508-11.
- Ghanghro AB, Lanjwani AH, Ghanghro IH, Khuhawar TM.

- Screening and quantitative analysis of phytochemicals from three selected medicinal plants. *Int J Pharm Med Res.* 2015;3(4):263-6.
28. Rastogi RP, Mehrotra BN. *Compendium of Indian Medicinal Plants. Vol I.* New Delhi: Publications & Information Directorate, Council of Scientific and Industrial Research; 1990. p. 28.
 29. Asolkar LV, Chopra RN. *Second Supplement to Glossary of Indian Medicinal Plants with Active Principles.* Publications & Information Directorate; 1992.
 30. Rastogi RP, Mehrotra BN. *Compendium of Indian Medicinal Plants. Vol III.* New Delhi, India: Central Drug Research Institute, Publications & Information Directorate; 1993.
 31. Daulatabad CD, Jamkhandi AM. *Althaea rosea* seed oil-a rich source of ricinoleic acid. *Journal of the Oil Technologists' Association of India.* 2000;32(1):8-9.
 32. Kwiatkowska M, Stepinski D, Poplonska K, Wojtczak A, Polit JT. 'Elaioplasts' identified as lipotubuloids in *Althaea rosea*, *Funkia sieboldiana* and *Vanilla planifolia* contain lipid bodies connected with microtubules. *Acta Soc Bot Pol.* 2011;80(3):211-9. doi: [10.5586/asbp.2011.036](https://doi.org/10.5586/asbp.2011.036).
 33. Dudek M, Matławska I, Szkudlarek MA. Phenolic acids in the flowers of *Althaea rosea* var. *nigra*. *Acta Pol Pharm.* 2006;63(3):207-11.
 34. Benbassat N, Yoncheva K, Hadjimitova V, Hristova N, Konstantinov S, Lambov N. Influence of the extraction solvent on antioxidant activity of *Althaea officinalis* L. root extracts. *Cent Eur J Biol.* 2014;9(2):182-8. doi: [10.2478/s11535-013-0245-2](https://doi.org/10.2478/s11535-013-0245-2).
 35. Ammar NM, El-Kashoury E, Abou El-Kassem LT, Abd El-Hakeem RE. Evaluation of the phenolic content and antioxidant potential of *Althaea rosea* cultivated in Egypt. *J Arab Soc Med Res.* 2013;8(2):48-52. doi: [10.4103/1687-4293.123786](https://doi.org/10.4103/1687-4293.123786).
 36. Nair AG, Nagarajan S, Subramanian SS. Flavonoids of the flowers of *Hibiscus abelmoschus* and *Althaea rosea*. *Cur Sci.* 1964;33(14):431-2.
 37. Khory RN, Katrak NN. *Materia Medica of India and Their Therapeutics.* Delhi: Neeraj Publishing House; 1981. p. 647.
 38. Khare CP. *Indian Medicinal Plants: An Illustrated Dictionary.* Delhi: Heidelberg Publication; 2007. p. 40.
 39. Abdel Salam NA, Ghazy NM, Shawky E, Sallam SM, Shenouda ML. Validated HPTLC method for dihydrokaempferol-4'-O-glucopyranoside quantitative determination in *Alcea* species. *J Chromatogr Sci.* 2018;56(6):518-23. doi: [10.1093/chromsci/bmy025](https://doi.org/10.1093/chromsci/bmy025).
 40. Blunden G, Patel AV, Armstrong NJ, Gorham J. Betaine distribution in the Malvaceae. *Phytochemistry.* 2001;58(3):451-4. doi: [10.1016/s0031-9422\(01\)00263-1](https://doi.org/10.1016/s0031-9422(01)00263-1).
 41. Hosaka H, Mizuno T, Iwashina T. Flavonoid pigments and color expression in the flowers of black hollyhock (*Alcea rosea* 'nigra'). *Bull Natl Mus Nat Sci Ser B.* 2012;38:69-75.
 42. Kohlmunzer S, Konska G, Wiatr E. Anthocyanosides of *Alcea rosea* L. var. *nigra* as vasoprotective agents. *Herb Hungarica.* 1983;22:13-22.
 43. Markham KR, Bloor SJ, Nicholson R, Rivera R, Shemluck M, Kevan PG, et al. Black flower coloration in wild *Lisianthus nigrescens*: its chemistry and ecological consequences. *Z Naturforsch C Biosci.* 2004;59(9-10):625-30. doi: [10.1515/znc-2004-9-1003](https://doi.org/10.1515/znc-2004-9-1003).
 44. Atkhamova SK, Rakhimova DA, Rakhmanberdyeva RK, Karimdzhanov AK, Ismailov AI. Plant polysaccharides II. A study of the carbohydrates of *Alcea rosea*. *Chem Nat Compd.* 1995;31(2):262-. doi: [10.1007/bf01170222](https://doi.org/10.1007/bf01170222).
 45. Classen B, Blaschek W. High molecular weight acidic polysaccharides from *Malva sylvestris* and *Alcea rosea*. *Planta Med.* 1998;64(7):640-4. doi: [10.1055/s-2006-957538](https://doi.org/10.1055/s-2006-957538).
 46. Rakhimov DA, Atkhamova SK, Khvan AM. Pectinic substances from *Alcea rosea* flowers. *Chem Nat Compd.* 2007;43(6):685-6. doi: [10.1007/s10600-007-0228-5](https://doi.org/10.1007/s10600-007-0228-5).
 47. Atkhamova SK, Rakhmanberdyeva RK, Rakhimov DA, Karimdzhanov AK, Ismailov AI, Khodzhimatov KK, et al. Carbohydrates of cultivated varieties of *Althaea rosea*. *Chem Nat Compd.* 1993;29(5):566-8. doi: [10.1007/bf00630197](https://doi.org/10.1007/bf00630197).
 48. Rastogi RP, Mehrotra BN. *Compendium of Indian Medicinal Plants. Vol II.* New Delhi: Publications & Information Directorate, Council of Scientific and Industrial Research; 1991. p. 37-8.
 49. Azab A. *Alcea*: traditional medicine, current research and future opportunities. *Eur Chem Bull.* 2016;5(12):505-14. doi: [10.17628/ecb.2016.5.505](https://doi.org/10.17628/ecb.2016.5.505).
 50. Jain SK. Ethnobotany and research on medicinal plants in India. In: Chadwick DJ, Marsh J, eds. *Ciba Foundation Symposium 185-Ethnobotany and the Search for New Drugs.* Chichester, UK: John Wiley & Sons; 2007. p. 153-68.
 51. Gairola S, Sharma J, Bedi YS. A cross-cultural analysis of Jammu, Kashmir and Ladakh (India) medicinal plant use. *J Ethnopharmacol.* 2014;155(2):925-86. doi: [10.1016/j.jep.2014.06.029](https://doi.org/10.1016/j.jep.2014.06.029).
 52. Nadkarni KM. *Indian Materia Medica. Vol I.* Bombay: Popular Prakashan; 1989.
 53. Tohmé G, Tohmé H. *A Thousand and One Flowers of Lebanon. Vol 22.* Lebanese University; 2002. p. 309.
 54. Biswas PK. *Encyclopaedia of Medicinal Plants. Vol 2.* New Delhi: Dominant Publishers & Distributors; 2006.
 55. Ahmadi M, Khajavi Rad A, Rajaei Z, Hadjzadeh M, Mohammadian N, Tabasi NS. *Alcea rosea* root extract as a preventive and curative agent in ethylene glycol-induced urolithiasis in rats. *Indian J Pharmacol.* 2012;44(3):304-7. doi: [10.4103/0253-7613.96298](https://doi.org/10.4103/0253-7613.96298).
 56. Munir M, Hussain A, Ul-Haq I, Qureshi R, Munazir M, Arshad M, et al. Callogenesis potential of cotyledonary explants of *Althaea rosea*. from Pakistan. *Pak J Bot.* 2012;44:271-5.
 57. Company ME. *Physicians' desk reference: Physician's Desk Reference (PDR).* 1999.
 58. Khan I, Abd-Ur-Rehman, Awan SA, Aslam S, Mursalin M, Shabbir M. Importance of ethnomedicinal flora of Sarai Alamgir (boundary side of river Jhelum) district Gujrat, Punjab, Pakistan. *Med Aromat Plants (Los Angel).* 2016;5(4):264. doi: [10.4172/2167-0412.1000264](https://doi.org/10.4172/2167-0412.1000264).
 59. Dar PA, Ali F, Sheikh IA, Ganie SA, Dar TA. Amelioration of hyperglycaemia and modulation of antioxidant status by *Alcea rosea* seeds in alloxan-induced diabetic rats. *Pharm Biol.* 2017;55(1):1849-55. doi: [10.1080/13880209.2017.1333127](https://doi.org/10.1080/13880209.2017.1333127).
 60. Pakravan M. *Alcea ilamica*, a new species from Iran. *Rostaniha.* 2003;4(2):123-4.
 61. Amiri MS, Joharchi MR. Ethnobotanical investigation of traditional medicinal plants commercialized in the markets of Mashhad, Iran. *Avicenna J Phytomed.* 2013;3(3):254-71.
 62. Ahmed I, Roy BC, Subramanian D, Ganie SA, Kwatra D, Dixon D, et al. An ornamental plant targets epigenetic signaling to block cancer stem cell-driven colon carcinogenesis. *Carcinogenesis.* 2016;37(4):385-96. doi: [10.1093/carcin/bgw009](https://doi.org/10.1093/carcin/bgw009).
 63. Ertas A, Boga M, Gazioglu I, Yesil Y, Hasimi N, Ozaşlan C, et al. Fatty acid, essential oil and phenolic compositions of *Alcea pallida* and *Alcea apterocarpa* with antioxidant, anticholinesterase and antimicrobial activities. *Chiang Mai J Science.* 2016;43(1):1143-53.
 64. Calderón-Montaño JM, Burgos-Morón E, Pérez-Guerrero C, López-Lázaro M. A review on the dietary flavonoid kaempferol. *Mini Rev Med Chem.* 2011;11(4):298-344. doi: [10.2174/138955711795305335](https://doi.org/10.2174/138955711795305335).
 65. Zhang Y, Jin L, Chen Q, Wu Z, Dong Y, Han L, et al. Hypoglycemic activity evaluation and chemical study on hollyhock flowers. *Fitoterapia.* 2015;102:7-14. doi: [10.1016/j.fitote.2015.02.001](https://doi.org/10.1016/j.fitote.2015.02.001).

66. Dongfeng W, Jiuyu S, Qinghai Y. Study on the effect of *Althaea rosea* on the cardiovascular system. 1988. Available from: http://en.cnki.com.cn/Article_en/CJFDTOTAL-SYYD198804008.
67. Hussain L, Akash MS, Tahir M, Rehman K, Ahmed KZ. Hepatoprotective effects of methanolic extract of *Alcea rosea* against acetaminophen-induced hepatotoxicity in mice. *Bangladesh J Pharmacol*. 2014;9(3):322-7. doi: 10.3329/bjp.v9i3.19068.
68. Berger PJ, Sanders EH, Gardner PD, Negus NC. Phenolic plant compounds functioning as reproductive inhibitors in *Microtus montanus*. *Science*. 1977;195(4278):575-7. doi: 10.1126/science.319531.
69. Papiez M, Gancarczyk M, Bilińska B. The compounds from the hollyhock extract (*Althaea rosea* Cav. var. *nigra*) affect the aromatization in rat testicular cells in vivo and in vitro. *Folia Histochem Cytobiol*. 2002;40(4):353-9.
70. Papiez MA. The influence of hollyhock extract administration on testicular function in rats. *J Mol Histol*. 2004;35(8-9):733-40. doi: 10.1007/s10735-004-5067-0.
71. Guinot P, Rogé A, Gargadennec A, Garcia M, Dupont D, Lecoœur E, et al. Dyeing plants screening: an approach to combine past heritage and present development. *Color Technol*. 2006;122(2):93-101. doi: 10.1111/j.1478-4408.2006.00015.x.
72. Lim TK. *Alcea rosea*. In: *Edible Medicinal and Non-Medicinal Plants*. Dordrecht: Springer; 2014.
73. Liu F, Du W, Bai X, Tian S. Quantification of phytochemical constituents and in-vitro antioxidant activity of *Althaea rosea* seeds. *J Chem Pharm Res*. 2014;6(3):1466-71.
74. Namjoyan F, Jahangiri A, Azemi ME, Arkian E, Mousavi H. Inhibitory effects of *Physalis alkekengi* L., *Alcea rosea* L., *Bunium persicum* B. Fedtsch. and *Marrubium vulgare* L. on mushroom tyrosinase. *Jundishapur J Nat Pharm Prod*. 2015;10(1):e23356. doi: 10.17795/jjnpp-23356.
75. Zheng W, Wang SY. Antioxidant activity and phenolic compounds in selected herbs. *J Agric Food Chem*. 2001;49(11):5165-70. doi: 10.1021/jf010697n.
76. Mert T, Fafal T, Kivçak B, Öztürk HT. Antimicrobial and cytotoxic activities of the extracts obtained from the flowers of *Alcea rosea* L. *Hacet Univ J Fac Pharm*. 2010;30(1):17-24.
77. Seyyednejad SM, Koochak H, Darabpour E, Motamedi H. A survey on *Hibiscus rosa-sinensis*, *Alcea rosea* L. and *Malva neglecta* Wallr as antibacterial agents. *Asian Pac J Trop Med*. 2010;3(5):351-5. doi: 10.1016/s1995-7645(10)60085-5.
78. Lee KH. Novel antitumor agents from higher plants. *Med Res Rev*. 1999;19(6):569-96. doi: 10.1002/(sici)1098-1128(199911)19:6 < 569::aid-med7 > 3.0.co;2-9.
79. Jiménez-Orozco FA, Molina-Guarneros JA, Mendoza-Patiño N, León-Cedeño F, Flores-Pérez B, Santos-Santos E, et al. Cytostatic activity of coumarin metabolites and derivatives in the B16-F10 murine melanoma cell line. *Melanoma Res*. 1999;9(3):243-7. doi: 10.1097/00008390-199906000-00005.
80. Matsumae R, Rani S, Motoki K. Preservation test of holly hock yam extract by xylon. *Echime Ken Sei Shi Shik Hoko*. 1956;9:9-13.

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