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Anti-SARS-CoV and Anti-cancer Effects of Emodin

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Abstract

Background and aims: The SARS-CoV-2 disease 2019 (COVID-19), whose spread started in the late December in 2019 in China, is the main concern in the world today. Potential anti-coronavirus targets can be categorized into two classes depending on the target, one is operating on the host immune system or human cells, and the other is on coronavirus itself. Anthraquinones are generally extracted from the Polygonaceae family, and have many beneficiary characteristics such as being antibacterial, anti-cancer and anti-diabetes. Emodin anthraquinones represent an important role in human health and have golden healthful features making them a drug to cure many illnesses. The aim of this study was to review the inhibiting effect of emodin on cancer and SARS-CoV-2.

Methods: This comprehensive literature review was performed on papers that have been published from 1994 till 2020 in various data resources such as NCBI, Science direct, Springer and Web of science. The selected keywords were emodin, medicinal plant, anticancer plant and medicinal herbs, cancer and SARS-CoV-2.

Results: Different studies were found that emodin is known as an effective agent to obstruct the interaction of the S protein of SARS-CoV and the host ACE2 (Angiotensin converting enzyme 2) and the infection caused by the retrovirus. In addition, the outbreak of cancer in patients infected by SARS-CoV-2 (COVID-19) is more than it among the general population.

Conclusion: Therefore, the present research is going to outline and highlight the anti SARS-CoV-2 therapeutic strategies of emodin and the anti-cancer characteristics' of this drug.

Keywords: Emodin, SARA-Cov-2, Cancer.

Review article

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Anti-SARS-CoV and Anti-cancer Effects of ...

Introcuction

The SARS-CoV-2 disease (COVID-19), whose breakout started in the late December in 2019 in Wuhan-China. is the main concern in the world today. It has had a quick breakout across China and then different countries 1,2 . The virus belongs to the β family and coronavirus evolved evolutionarily by bats, and then it was transferred to human beings ^{3,4}. The genome sequence of SARS-CoV-2 shows 96.2% similarity with SARSrelated coronavirus (SARSr-CoV; RaTG13), 79% with SARS-CoV, and 50% with MERS-CoV ^{4,5}. SARS-CoVreceptor is angiotensin 2'sthe converting enzyme II (ACE2) and uses a spike protein as an attachment to its receptor ⁶⁻⁸. It is believed that partial spike gene of novel corona virus is produced by a pangolin type⁹⁻¹² coronavirus. SARS-CoV-2 contains a positive RNA genome, and it has at least four structural proteins: Spike (S) protein. protein. envelope (E) (M) protein, membrane and (N) protein^{12,13}. The nucleocapsid results of several recent studies have represented a correlation between and COVID-19¹⁴. cancer Patients suffering from cancer have weaker immune systems in comparison with the general population both due to the nature of the disease and its treatment processes. The rates of infection and mortality are high among cancer patients. Research shows that the outbreak of cancer has been more among patients infected by SARS-CoV-

2 (COVID-19) than among the general population ¹⁵. In China, it is known that 1% of the patients infected to COVID-19 have had a history of cancer. Lung cancer is the most common cancer among these patients while colorectal, breast, and bladder cancer, lymphoma, papillary thyroid cancer, renal cell carcinoma, adrenal carcinoma are in the forthcoming ranks ¹⁵.

Potential anti-coronavirus therapies

Potential anti-coronavirus targets can be categorized into two classes depending on the target, one is operating on the host immune system or human cells, and the other is on coronavirus itself ¹⁶. Regarding human targets, the SARS-CoV-2, similar to SARS virus, binds to the angiotensin-converting enzyme 2 (ACE2) receptor which is expressed abundantly in lung, kidney, heart and some other organs ^{17,18}. The virus uses transmembrane protease, serine2 (TMPRSS2) in order to spike protein activation. Spike is broken into S1 and S2 by TMPRSS2. This plays a crucial role in SARS and coronavirus infection:therefore, TMPRSS2 could be a novel antiviral strategy against coronavirus and some influenza viruses ¹⁹⁻²⁷.

Drug Targets for Coronavirus

The pharmacological targets of coronavirus are as follows:

1-Papain-like proteinase (PLpro) is responsible for the production of Nsp1, Nsp2 and Nsp3 through an N-terminus breakage of the replicase poly-protein. These proteins are important for virus

genome replication ²⁸. PLpro works as an antagonism of the host's innate immunity ²⁹⁻³¹.

2-3C-like main protease (3CLpro or Nsp5) is another therapeutic target for the novel coronavirus. This enzyme is essentially maturates itself using polyproteins and then produces an Nsp4–Nsp16³².

3-RNA-dependent RNA polymerase (RdRp or Nsp12) is a very important protein in novel virus structure which contains Ser-Asp-Asp motif in an active site, and its activity is enhanced through NSP12 ^{33, 34}.

4- Helicase (Nsp13) is utilized for separating double-stranded (ds) DNA and RNA and contains two domains: zinc binding domain located on N-terminal, and a helicase domain ³⁵.

5-Some non-structural proteins which are involved in virus RNA synthesis can be targets for drug design. In this regard, NSP-3b, 3e, 7, 9. 10, 14, 15 and NSP7-8 complexes are candidates of the virus inhibition¹⁶. Structural proteins: Spike is the vital structural protein which is important in virus interaction to host cell receptors ³⁶. Spike protein is broken into S1 and S2 by the TMPRSS2 which corporate in attachment and fusion of virus to host cell ³⁷.

6- Virulence factors of coronavirus: Nsp1, Nsp3c and ORF7a are recognized as virulence factors. Nsp1 leads to the inhibition of producing type-I interferon and degradation of mRNA by attaching to host 40S ribosomal subunit ^{38,39}. Nsp3c is involved in resistance of the novel virus to host innate immunity. The process occurs via attachment of this virus to host ADP-ribose ⁴⁰. Bone marrow stromal antigen 2 (BST-2 or

tetherin) is a pre-B-cell growth activator $\overset{41,42}{,},$ and it is a marker of type I cells $(IPC)^{43}$. interferon-producing BST-2 has an antiviral activity which diminishes the release of human coronavirus 229E (hCoV-229E) and many other viruses ⁴⁴. Antiviral activity of BST-2 is restricted by ORF7a via direct interaction of ORF7a with BST-2 ⁴⁵. It has been shown that SARS-CoV and other coronaviruses have an open reading frame ORF-3a that encodes a monovalent cations-permeable channel in the infected host cells. The activity of this channel affects virus release and it entails a higher selectivity of K+ compared to Na+ 46-48.

Emodin and its beneficiary effects on Human Health

Anthraquinones are generally Polygonaceae extracted from the family, such as Rheum palmatum and Rheum officinale. Anthraquinones have many beneficiary features such as being antibacterial, anti-cancer, antidiabetes ^{49,50}. Emodin anthraquinones (1,3,8-trihydroxy-6-ethylanthraquinone) (Fig-1) plays important roles in human health and it entails golden healthful features making it a drug for the treatment of the following illnesses: gallstones, inflammation and inflammatory diseases, hepatitis ^{51,52}. Emodin has been utilized as a laxative therapy for many years and some laxative mechanisms of emodin are as follows: the reduction of Na+-K+-ATPase activity in intestinal mucosa, inhibition of somatostatin, and enhancing the release of acetyl choline ⁵²-⁵⁴



Figure1- structure of emodin (55)

It has been shown that emodin is an anti-inflammatory, anti-ulcerogenic, anticancer, immunosuppressive, antibacterial, antiviral ^{56,57}, vasorelaxant, and a chemo preventive factor 58 .

Anti-cancer effects of emodin

Emodin induces apoptosis in a dosedependent manner, and activates caspase-3 ⁵⁹ and -9 enzymes ⁶⁰, induces p53 protein ⁶¹, generates reactive oxygen species (ROS) ⁶², downregulates androgen receptors ⁶³, suppresses lipid raft coalescence ⁶⁴, inhibits Janusactivated kinase 2 ⁶⁵, and it utilizes some apoptotic and anticancer mechanisms in various cell types (Fig-2).



Figure2- Anti-cancer mechanisms of emodin

The Vital Role of Emodin in Novel Coronavirus Treatment

Various studies have demonstrated four drug targets of Emodin in inhibiting the coronavirus. This compound is a potent inhibitor of the 3a ion channel (Fig-3). Inhibition of this channel can counteract the release of the virus. The reduction of extracellular viral RNA copied by emodin is an evidence of inhibition of the virus release. This drug leads to decrease intracellular scripts of the virus 'RNA' in the presence of high concentrations. This indicates that emodin can inhibit other stages of the virus' life cycle at higher concentrations ^{66, 67}. Another

mechanism of emodin in inhibiting the coronavirus is approved regarding SARS-Cov by blocking the virus binding to its receptor, the angiotensin-converting enzyme ⁶⁸. The binding of the S protein to ACE2 is inhibited in 200 μ M of emodin and 80% inhibition is achieved in 50 μ M dosage ⁶⁵. Also emodin is a strong inhibitor of the 3a channel in about 20 M ⁶⁶.

Still another emodin treatment strategy is inhibition of the 3C like protease (3CLpro) enzyme in novel virus which automatically cleaves polyproteins produce to mature enzymes, and its cleavage site is the downstream area of the non-structural protein leading to release the nonstructural protein NSP4-NSP16s. As a result, this enzyme is involved in the maturation of non-structural proteins and therefore; it is considered as an essential enzyme for the viral life cycle to be utilized as a new drug target 69 .

It has been shown that when 3CLpro enzyme is inhibited, the replication of the SARS-cov virus in the host cell is inhibited 70, and emodin leads to the inhibition of this enzyme which is present in Covid-19 virus structure 69-71. It has been shown that emodin Aloe (1,8-dihydroxy-3-(hydroxymethyl) anthraquinone) is a another type of emodin which is present in aloe latex, in dose-dependently manner inhibited cleavage activity of the SARS coronavirus 3CLpro, in cellfree (the IC50 values were 132µM), and in cell-based assays ⁷².

Emodin inhibit the Janus-activated kinase-2 enzyme and the JAK2 / STAT3 signaling pathways induced by interleukin-6⁶⁵. Furthermore, it leads to the inhibition of the cytokine storm because the release of interleukin-6 releases other cytokines, resulting in inflammatory storms and the death of the infected patients⁷³. It is worth mentioning that there are at least 36 IL-6 inhibitors which only two of them are monoclonal antibodies: Tocilizumab and Sarilumab, approved by the FAD. It has been shown that dietary Emodin has antioxidant properties and reduces oxidative damage to organs ⁷⁴.

Side effects of emodin:

Emodin also has some side effects, if it is consumed in high dosages for a long period of time. In this regard, genotoxic effects of emodin are reported in some studies⁷⁵. Furthermore, Emodin is carcinogen in rodents, and leads to cancer in some animals. Others studies the results of the have rejected emodin⁷⁶-⁷⁹. of carcinogenicity However, evaluating the genotoxity profile of emodin did not show any concerns on its genotoxity in humans⁷⁹. Because emodin is a potent inhibitor of cytochrome P4501A1, and most prooncogen compounds need to be activated by detoxifying enzymes, emodin has been shown to inhibit this protein and thus counteracts the mutating effects of P4501A1. According to the above sources⁸⁰, the lack of genotoxicity of emodin in humans is supported.



Figure 3 - Anti SARS-CoV -2 mechanisms of emodin

Conclusion

Emodin may operate as an antiviral drug by obstructing virus infection and its release. In addition, it shows anticancer effects using various mechanisms. Hence, emodin may open the horizons to novel therapeutics in treatment of coronaviruses. It can be considered as basis for а drug development coronavirus against infections and various cancers.

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Conflict of Interest

All authors report no conflicts of interest relevant to this article.

References

1. Cui J, Li F, Shi Z-L. Origin and evolution of pathogenic coronaviruses. *Nature Reviews Microbiology*. 2019; 17(3): 181-92.

2. Li X, Song Y, Wong G, Cui J. Bat origin of a new human coronavirus:

there and back again. Science China Life Sciences. 2020; 63(3): 461-2.

3. Li W, Shi Z, Yu M, Ren W, Smith C, Epstein JH, et al. 2191341. Bats Are Natural Reservoirs of SARS-Like Coronaviruses. *Science*. 2005; 310(5748): 676-9.

4. Dominguez SR, O'Shea TJ, Oko LM, Holmes KV. Detection of group 1 coronaviruses in bats in North America. *Emerging Infectious Diseases*. 2007; 13(9): 1295.

5. Gralinski LE, Menachery VD. Return of the Coronavirus: 2019-nCoV. *Viruses.* 2020; 12(2): 135.

6. Zhou P, Yang X-L, Wang X-G, Hu B, Zhang L, Zhang W, et al. Discovery of a novel coronavirus associated with the recent pneumonia outbreak in humans and its potential bat origin. *Bio Rxiv*. 2020.

7. Li X, Song Y, Wong G, Cui J. Bat origin of a new human coronavirus: there and back again. *Science China Life Sciences* . 2020; 63(3): 461-2.

8. Wei X, Li X, Cui J. Evolutionary perspectives on novel coronaviruses identified in pneumonia cases in China.

National Science Review. 2020; 7(2): 239-42.

9. Wong MC, Cregeen SJJ, Ajami NJ, Petrosino JF. Evidence of recombination in coronaviruses implicating pangolin origins of nCoV-2019. *Biorxiv*. 2020.

10. Xiao K, Zhai J, Feng Y, Zhou N, Zhang X, Zou J-J, et al. Isolation and characterization of 2019-nCoV-like coronavirus from Malayan pangolins. *BioRxiv*. 2020.

11. Lam TT-Y, Shum MH-H, Zhu H-C, Tong Y-G, Ni X-B, Liao Y-S, et al. Identification of 2019-nCoV related coronaviruses in Malayan pangolins in southern China. *BioRxiv*. 2020.

12. Tang X, Wu C, Li X, Song Y, Yao X, Wu X, et al. On the origin and continuing evolution of SARS-CoV-2. *National Science Review*. 2020.

13. Bosch BJ, van der Zee R, de Haan CA, Rottier PJ. The coronavirus spike protein is a class I virus fusion protein: structural and functional characterization of the fusion core complex. *Journal of Virology*. 2003; 77(16): 8801-11.

14. Cancarevic I, Tathineni P, Malik
BH. Coronavirus Disease 2019
(COVID-19) in Cancer Patients.
Cureus. 2020; 12(4): e7835.

15. Liang W, Guan W, Chen R, Wang W, Li J, Xu K, et al. Cancer patients in SARS-CoV-2 infection: a nationwide analysis in China. The Lancet Oncology. 2020; 21(3): 335-7.

16. Wu C, Liu Y, Yang Y, Zhang P, Zhong W, Wang Y, et al. Analysis of therapeutic targets for SARS-CoV-2 and discovery of potential drugs by computational methods. Acta Pharmaceutica Sinica B. 2020.

17. Wang C, Horby PW, Hayden FG, Gao GF. A novel coronavirus outbreak of global health concern. *The Lancet*. 2020; 395(10223): 470-3.

18. Zhao Y, Zhao Z, Wang Y, Zhou Y, Ma Y, Zuo W. Single-cell RNA expression profiling of ACE2, the putative receptor of Wuhan 2019-nCov. *BioRxiv*. 2020.

19. Sakai K, Ami Y, Tahara M, Kubota T, Anraku M, Abe M, et al. The host protease TMPRSS2 plays a major role in in vivo replication of emerging H7N9 and seasonal influenza viruses. *Journal of Virology*. 2014; 88(10): 5608-16.

20. Tarnow C, Engels G, Arendt A, Schwalm F, Sediri H, Preuss A, et al. TMPRSS2 is a host factor that is essential for pneumotropism and pathogenicity of H7N9 influenza A virus in mice. *Journal of virology*. 2014; 88(9): 4744-51.

21. Matsuyama S, Nagata N, Shirato K, Kawase M, Takeda M, Taguchi F. Efficient activation of the severe acute respiratory syndrome coronavirus spike protein by the transmembrane protease TMPRSS2. *Journal of Virology*. 2010; 84(24): 12658-64.

22. Shirato K, Kawase M, Matsuyama S. Middle East respiratory syndrome coronavirus infection mediated by the transmembrane serine protease TMPRSS2. *Journal of Virology*. 2013; 87(23): 12552-61.

23. Böttcher-Friebertshäuser E, Klenk H-D, Garten W. Activation of influenza viruses by proteases from host cells and bacteria in the human airway epithelium. *Pathogens diseases*. 2013; 69(2): 87-100.

24. Momattin H, Mohammed K, Zumla A, Memish ZA, Al-Tawfiq JA. Therapeutic options for Middle East respiratory syndrome coronavirus (MERS-CoV)-possible lessons from a review of SARS-CoV systematic therapy. International Journal of Infectious Diseases. 2013; 17(10): е792-е8.

25. Zumla A, Chan JF, Azhar EI, Hui DS, Yuen K-Y. Coronaviruses—drug discovery and therapeutic options. Nature reviews Drug discovery. 2016; 15(5): 327.

26. Yamaya M, Shimotai Y, Hatachi Y, Homma M, Nishimura H. Serine Proteases and their Inhibitors in Human Airway Epithelial. 2016.

27. Cheng Z, Zhou J, To KK-W, Chu H, Li C, Wang D, et al. Identification of TMPRSS2 as a susceptibility gene for severe 2009 pandemic A (H1N1) influenza and A (H7N9) influenza. The Journal of Infectious Diseases. 2015; 212(8): 1214-21.

28. Harcourt BH, Jukneliene D, Kanjanahaluethai A, Bechill J, Severson KM, Smith CM, et al. Identification of severe acute respiratory syndrome coronavirus replicase products and characterization of papain-like protease activity. *Journal of Virology*. 2004; 78(24): 13600-12.

29. Chen X, Yang X, Zheng Y, Yang Y, Xing Y, Chen ZJP. SARS coronavirus papain-like protease inhibits the type I interferon signaling pathway through interaction with the STING-TRAF3-TBK1 complex. *Protein Cell.* 2014; 5(5): 369-81.

30. Yuan L, Chen Z, Song S, Wang S, Tian C, Xing G, et al. p53 degradation by a coronavirus papain-like protease suppresses type I interferon signaling. *Journal of Biological Chemistry*. 2015; 290(5): 3172-82.

31. Li S-W, Wang C-Y, Jou Y-J, Huang S-H, Hsiao L-H, Wan L, et al. SARS coronavirus papain-like protease inhibits the TLR7 signaling pathway Lys63-linked removing through polyubiquitination of TRAF3 and TRAF6. International Journal of Molecular Sciences. 2016; 17(5): 678.

32. Yang H, Xie W, Xue X, Yang K, Ma J, Liang W, et al. Design of widespectrum inhibitors targeting coronavirus main proteases. *PLoS Biology*. 2005; 3(10): e324.

33. Subissi L, Imbert I, Ferron F, Collet A, Coutard B, Decroly E, et al.
SARS-CoV ORF1b-encoded nonstructural proteins 12–16: replicative enzymes as antiviral targets.
Antiviral Research. 2014; 101: 122-30.
34. Imbert I, Guillemot JC, Bourhis JM, Bussetta C, Coutard B, Egloff MP, et al. A second, non- canonical RNAdependent RNA polymerase in SARS Coronavirus. The EMBO Journal.

2006; 25(20): 4933-42.

35. Ivanov KA, Ziebuhr J. Human coronavirus 229E nonstructural protein 13: characterization of duplexunwinding, nucleoside triphosphatase, and RNA 5'-triphosphatase activities. *Journal of Virology*. 2004; 78(14): 7833-8.

36. Millet JK, Whittaker GR. Host cell proteases: Critical determinants of coronavirus tropism and pathogenesis. *Virus Research.* 2015; 202: 120-34.

37. Xia S, Liu Q, Wang Q, Sun Z, Su S, Du L, et al. Middle East respiratory syndrome coronavirus (MERS-CoV) entry inhibitors targeting spike protein. *Virus Research.* 2014; 194: 200-10.

38. Kamitani W, Narayanan K, Huang C, Lokugamage K, Ikegami T, Ito N, et al. Severe acute respiratory syndrome coronavirus nsp1 protein suppresses host gene expression by promoting host mRNA degradation. *Proceedings of the National Academy of Sciences*. 2006; 103(34): 12885-90.

39. Narayanan K, Huang C, Lokugamage K, Kamitani W, Ikegami T, Tseng C-TK, et al. Severe acute respiratory syndrome coronavirus nsp1 suppresses host gene expression, including that of type I interferon, in infected cells. *Journal of Virology*. 2008; 82(9): 4471-9.

40. Forni D, Cagliani R, Mozzi A, Pozzoli U, Al-Daghri N, Clerici M, et al. Extensive positive selection drives the evolution of nonstructural proteins in lineage C betacoronaviruses. *Journal of Virology*. 2016; 90(7): 3627-39.

41. Goto T, Kennel SJ, Abe M, Takishita M, Kosaka M, Solomon A, et al. A novel membrane antigen selectively expressed on terminally differentiated human B cells. 1994.

42. Ishikawa J, Kaisho T, Tomizawa H, Lee BO, Kobune Y, Inazawa J, et al. Molecular cloning and chromosomal mapping of a bone marrow stromal cell surface gene, BST2, that may be involved in pre-B-cell growth. *Genomics.* 1995; 26(3): 527-34.

43. Blasius AL, Giurisato E, Cella M, Schreiber RD, Shaw AS, Colonna M. Bone marrow stromal cell antigen 2 is a specific marker of type I IFN-producing cells in the naive mouse, but a promiscuous cell surface antigen following IFN stimulation. *The Journal of Immunology*. 2006; 177(5): 3260-5.

44. Taylor JK, Coleman CM, Postel S, Sisk JM, Bernbaum JG, Venkataraman T, et al. Severe acute respiratory syndrome coronavirus ORF7a inhibits bone marrow stromal antigen 2 virion tethering through a novel mechanism of glycosylation interference. *Journal of Virology*. 2015; 89(23): 11820-33.

45. Taylor JK, Coleman CM, Postel S, Sisk JM, Bernbaum JG, Venkataraman T, et al. Severe acute respiratory syndrome coronavirus ORF7a inhibits bone marrow stromal antigen 2 virion tethering through a novel mechanism of glycosylation interference. *Journal of Virology*. 2015; 89(23): 11820-33.

46. Zeng R, Yang R-F, Shi M-D, Jiang M-R, Xie Y-H, Ruan H-Q, et al. Characterization of the 3a protein of SARS-associated coronavirus in infected vero E6 cells and SARS patients. *Journal of molecular biology*. 2004; 341(1): 271-9.

47. Wang K, Xie S, Sun BJBeBA-B.Viral proteins function as ion channels.2011; 1808(2): 510-5.

48. Lu W, Zheng B-J, Xu K, Schwarz W, Du L, Wong CK, et al. Severe acute respiratory syndrome-associated coronavirus 3a protein forms an ion channel and modulates virus release. Proceedings of the National Academy of Sciences. 2006; 103(33): 12540-5.

49. Peng W, Qin R, Li X, Zhou H. Botany, phytochemistry, pharmacology, and potential application of Polygonum cuspidatum Sieb. et Zucc.: a review. Journal of ethnopharmacology. 2013; 148(3): 729-45.

50. Shan B, Cai Y-Z, Brooks JD, Corke HJFC. Antibacterial properties of Polygonum cuspidatum roots and their major bioactive constituents. 2008; 109(3): 530-7.

51. Dong X, Fu J, Yin X, Cao S, Li X, Lin L, et al. Emodin: a review of its pharmacology, toxicity and pharmacokinetics. *Phytotherapy Research.* 2016; 30(8): 1207-18.

52. Srinivas G, Babykutty S, Sathiadevan PP, Srinivas P. Molecular mechanism of emodin action: transition from laxative ingredient to an antitumor agent. *Medicinal research reviews*. 2007; 27(5): 591-608.

53. Zhang H-Q, Zhou C-H, Wu Y-Q. Effect of emodin on small intestinal peristalsis of mice and relevant mechanism. World Journal of Gastroenterology: WJG. 2005; 11(20): 3147.

54. Ali S, Watson M, Osborne R. The stimulant cathartic, emodin, contracts the rat isolated ileum by triggering release of endogenous acetylcholine. Autonomic Autacoid Pharmacology. 2004; 24(4): 103-5.

55. Hsu S-C, Chung J-G. Anticancer potential of emodin. *BioMedicine*. 2012; 2(3): 108-16.

56. Teng Z-h, Zhou S-y, Ran Y-h, Liu X-y, Yang R-t, Yang X, et al. Cellular absorption of anthraquinones emodin and chrysophanol in human intestinal Caco-2 cells. Bioscience, biotechnology, biochemistry. 2007; 71(7): 1636-43.

57. Lee H-Z, Hsu S-L, Liu M-C, Wu C-H. Effects and mechanisms of aloeemodin on cell death in human lung squamous cell carcinoma. *European Journal of pharmacology*. 2001; 431(3): 287-95.

58. Koyama J, Morita I, Tagahara K, Nobukuni Y, Mukainaka T, Kuchide M, et al. Chemopreventive effects of emodin and cassiamin B in mouse skin carcinogenesis. *Cancer letters*. 2002; 182(2): 135-9.

59. Chen Y-C, Shen S-C, Lee W-R, Hsu F-L, Lin H-Y, Ko C-H, et al. Emodin induces apoptosis in human promyeloleukemic HL-60 cells accompanied by activation of caspase 3 cascade but independent of reactive oxygen species production. Biochemical pharmacology. 2002; 64(12): 1713-24.

60. Srinivas G, Anto RJ, Srinivas P, Vidhyalakshmi S, Senan VP, Karunagaran D. Emodin induces apoptosis of human cervical cancer cells through poly (ADP-ribose) polymerase cleavage and activation of caspase-9. European Journal of pharmacology. 2003; 473(2-3): 117-25.

61. Shieh D-E, Chen Y-Y, Yen M-H, Chiang L-C, Lin C-C. Emodin-induced apoptosis through p53-dependent pathway in human hepatoma cells. *Life Sciences*. 2004; 74(18): 2279-90.

62. Yi J, Yang J, He R, Gao F, Sang H, Tang X, et al. Emodin enhances arsenic trioxide-induced apoptosis via generation of reactive oxygen species and inhibition of survival signaling. *Cancer Research*. 2004; 64(1): 108-16.

63. Cha T-L, Qiu L, Chen C-T, Wen Y, Hung M-C. Emodin down-regulates androgen receptor and inhibits prostate cancer cell growth. *Cancer Research*. 2005; 65(6): 2287-95.

64. Huang Q, Shen H-M, Shui G, Wenk MR, Ong C-N. Emodin inhibits tumor cell adhesion through disruption of the membrane lipid Raft-associated integrin signaling pathway. *Cancer Research.* 2006; 66(11): 5807-15.

65. Muto A, Hori M, Sasaki Y, Saitoh A, Yasuda I, Maekawa T, et al. Emodin has a cytotoxic activity against human multiple myeloma as a Janus-activated kinase 2 inhibitor. Molecular Cancer Therapeutics. 2007; 6(3): 987-94.

66. Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. The Lancet. 2020; 395(10224): 565-74.

67. Schwarz S, Wang K, Yu W, Sun B, Schwarz W. Emodin inhibits current through SARS-associated coronavirus 3a protein. *Antiviral Research*. 2011; 90(1): 64-9.

68. Luo W, Su X, Gong S, Qin Y, Liu W, Li J, et al. Anti-SARS coronavirus 3C-like protease effects of Rheum palmatum L. extracts. *Bioscience Trends*. 2009; 3(4): 6-124.

69. Pillaiyar T. Manickam M. Namasivayam V, Hayashi Y, Jung S-H. An Overview of Severe Acute Respiratory Syndrome-Coronavirus (SARS-CoV) 3CL Protease Inhibitors: Peptidomimetics and Small Molecule Chemotherapy. Journal of Medicinal Chemistry. 2016; 59(14): 6595-628.

70. Piccolella S, Cresente G, Faramarzi S, FFormato M, Tommasino Pecoraro M, Pacifico S. Polyphenols vs. Coronaviruses: How Far Has Research Moved Forward?. *Molecules*. 2020; 25(18): 4103. 71. Yang H, Yang M, Ding Y, Liu Y, Lou Z, Zhou Z, et al. The crystal structures of severe acute respiratory syndrome virus main protease and its complex with an inhibitor. *Proceedings of the National Academy of Sciences*. 2003; 100(23): 13190-5.

72. Lin C-W, Tsai F-J, Tsai C-H, Lai C-C, Wan L, Ho T-Y, et al. Anti-SARS coronavirus 3C-like protease effects of Isatis indigotica root and plant-derived phenolic compounds. 2005; 68(1): 36-42.

73. Masjedi A, Hashemi V, Hojjat-Farsangi M, Ghalamfarsa G, Azizi G, Yousefi M, et al. The significant role of interleukin-6 and its signaling pathway in the immunopathogenesis and treatment of breast cancer. *Biomedicine Pharmacotherapy*. 2018; 108: 1415-24.

74. Masjedi A, Hashemi V, Hojjat-Farsangi M, Ghalamfarsa G, Azizi G, Yousefi M, et al. The significant role of interleukin-6 and its signaling pathway in the immunopathogenesis and treatment of breast cancer. *Biomedicine Pharmacotherapy*. 2018; 108: 1415-24.

75. Liang Z, Chen H, Yu Z, Zhao Z. Comparison of raw and processed Radix Polygoni Multiflori (Heshouwu) by high performance liquid chromatography and mass spectrometry. *Chinese Medicine*. 2010; 5(1): 29.

76. Stark A, Townsend J, Wogan G, Demain A, Manmade A, Ghosh A. Mutagenicity and antibacterial activity of mycotoxins produced by Penicillium islandicum Sopp and Penicillium rugulosum. *Journal of Environmental Pathology Toxicology Letters*. 1978; 2(2): 313-24. 77. Van der Hoeven J, editor Occurrence and detection of natural mutagens and modifying factors in food products. Princess Takamatsu Symposia; 1985.

78. Bruggeman I, Van der Hoeven J. Lack of activity of the bacterial mutagen emodin in HGPRT and SCE assay with V79 Chinese hamster cells. *Mutation Research/Genetic Toxicology*. 1984; 138(2-3): 219-24. 79. Mengs U, Krumbiegel G, Völkner
W. Lack of emodin genotoxicity in the mouse micronucleus assay. Mutation
Research/Genetic Toxicology
Environmental Mutagenesis. 1997;
393(3): 289-93.

80. Brusick D, Mengs U. Assessment of the genotoxic risk from laxative senna products. Environmental Molecular Mutagenesis. 1997; 29(1): 1-9.